Antennal Lobe Processing Increases Separability of Odor Mixture Representations in the Honeybee

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Deisig N, Giurfa M, Sandoz JC. Antennal lobe processing increases separability of odor mixture representations in the honeybee. J Neurophysiol 103: 2185–2194, 2010. First published February 24, 2010; doi:10.1152/jn.00342.2009. Local networks within the primary olfactory centers reformat odor representations from olfactory receptor neurons to second-order neurons. By studying the rules underlying mixture representation at the input to the antennal lobe (AL), the primary olfactory center of the insect brain, we recently found that mixture representation follows a strict elemental rule in honeybees: the more a component activates the AL when presented alone, the more it is represented in a mixture. We now studied mixture representation at the output of the AL by imaging a population of second-order neurons, which convey AL processed odor information to higher brain centers. We systematically measured odor-evoked activity in 22 identified glomeruli in response to four single odors and all their possible binary, ternary, and quaternary mixtures. By comparing input and output responses, we determined how the AL network reformats mixture representation and what advantage this confers for odor discrimination. We show that increased inhibition within the AL leads to more synthetic, less elemental, mixture representation at the output level than that at the input level. As a result, mixture representations become more separable in the olfactory space, thus allowing better differentiation among floral blends in nature.

INTRODUCTION

The architecture of central primary olfactory centers is similar in vertebrates and insects. Both the vertebrate olfactory bulb (OB) and the insect antennal lobe (AL) consist of glomeruli, which receive input from olfactory receptor neurons (ORNs) expressing the same receptor type (Mombaerts et al. 1996; Vosshall et al. 2000). Glomeruli are interconnected by different sets of local neurons, which are mostly inhibitory and reformat odor representations in the form of a stimulus-dependent, spatiotemporal redistribution of activity across the AL/OB. The processed signals are further conveyed to higher-order centers by output neurons, the projection neurons (PNs) in insects, and the mitral/tufted cells in vertebrates (Fig. 1A; Bandawat et al. 2007; Laurent 2002; Root et al. 2007; Sachse and Galizia 2002; Wachowiak and Shipley 2006).

In Drosophila, studies on odor coding using single odor molecules have shown nonlinear signal transformation in the AL (Bhandawat et al. 2007; Wilson et al. 2004), which seems to increase separability among odor representations. However, whether such a principle applies to odor mixtures remains unknown because most studies on the neural reformatting of odor representations have essentially used single odorants (Bhandawat et al. 2007; Ng et al. 2002; Wang et al. 2003; Wilson et al. 2004). Because natural odors are complex blends including many different components, studying how the neural code for olfactory mixtures is reshaped by AL processing is imperative to understand odor processing in a natural framework. At the OB/AL input level, mixture representation follows essentially elemental rules because it can be predicted from the responses to the components (Carlsson et al. 2007; Deisig et al. 2006; Silbering and Galizia 2007; Tabor et al. 2004). By contrast, at the AL output level, strong interactions between component signals within the AL networks were found in the zebrafish (Tabor et al. 2004) and Drosophila (Silbering and Galizia 2007), assigning unique properties to each mixture’s representation and making it different from the sum of its components. It is still unclear whether such nonlinear interactions are a by-product of local processing or whether they provide an actual benefit for mixture discrimination.

Using in vivo optical imaging in honeybees, we previously recorded the calcium responses to four single odors and all their possible mixture combinations in a set of well-identified olfactory glomeruli (Deisig et al. 2006). By bath-applying the calcium-sensitive dye Calcium Green-2 AM on the brain, we recorded a compound signal dominated by sensory input (see DISCUSSION). In the present study, to unravel the transformation of mixture representation within the AL network, we backfilled a population of PNs with the calcium-sensitive dye fura-2 dextran and recorded the calcium responses of the same glomeruli to the same four single odors and all their possible mixture combinations. Thus input and output mixture representations could be compared. We show that PN mixture representation is the result of subtle reformatting within the AL, which makes similarity relationships between mixture and components less predictable based on component information, i.e., less elemental. Furthermore, we show for the first time that such nonelemental reformatting increases separability among odor mixtures, thus facilitating olfactory mixture discrimination.

METHODS

Preparation and staining

Worker honey bees (Apis mellifera) were caught at the entrance of outdoor hives, cooled on ice, and mounted in Plexiglas chambers. The antennae were oriented to the front and their base was fixed with two-component silicon (KwikSil; World Precision Instruments, Sarasota, FL), providing a seal between the flagella (remaining in the air for odor stimulation) and the brain (under Ringer solution for optical imaging). The head capsule was then opened and all glands, membranes, and trachea were removed to reveal the two antennal lobes.
In vivo calcium imaging recordings were done using an epifluorescent microscope (Olympus BX-51WI) with a $20 \times$ NA 0.5 water immersion objective (Olympus UMPPlanFL) and imaging system (T.I.L.L. Photonics, Gräfeling, Germany). Images were taken using a 640 $\times$ 480 pixel 12-bit monochrome charge-coupled device camera (T.I.L.L. Imago) cooled to $-12^\circ$C. The preparation was alternately excited with 340 and 380 nm monochromatic light (T.I.L.L. PolyChrom IV). Each recording consisted of 100 double frames, at a rate of 5 double frames/s. We used $4 \times 4$ binning on chip (pixel size corresponds to $\sim 2 \times 2 \mu$m). Integration time was 16–32 and 4–8 ms, respectively, for 340 and 380 nm excitation. Odor stimuli were applied just before the beginning of the 15th double frame until the 20th double frame (i.e., odor lasted 1 s) using a computer-driven odor supplying device as in Deisig et al. (2006). The primary odors were 1-hexanol, 2-octanol, linalool, and limonene (Sigma-Aldrich), each subjected to up to three runs of 17 stimulations, each including the four primary odors (A, B, C, and D), their binary (AB, AC, AD, BC, BD, and CD), ternary (ABC, ACD, BCD, and AB), and quaternary (ABCD) mixtures as well as air and 1-nonanol, included as controls. The brain was regularly washed with Ringer (in mM: NaCl, 130; KCl, 6; CaCl$_2$, 5; MgCl$_2$, 4; sucrose, 160; glucose, 25; HEPES, 10; pH 6.7, 500 mOsmol; all chemicals were from Sigma-Aldrich, Lyon, France) and before staining, illumination was reduced and switched to red light. In this work we specifically stained according to the protocol proposed by Sachse and Galizia (2002) projection neurons conveying odor information through the lateral antennocerebral tract (LACT) from the AL to the mushroom bodies and the lateral horn using fura-2 dextran (10,000 MW, in 2% BSA; Molecular Probes). After staining, the brain was thoroughly washed with saline and the bee was left in a dark place for 3 h.

**Optical recordings of odor-evoked activity**

Fluorescence photographs allowed visualization and identification of 22 glomeruli in all preparations, based on the standardized AL atlas (Galizia et al. 1999). From these, 21 were imaged in a previous study (Deisig et al. 2006) and were used for comparisons between processing levels. This population of imaged glomeruli has proved to account for olfactory behavioral performances of bees (Guerrieri et al. 2005).

**Raw data processing**

Analysis of calcium-imaging data was carried out using IDL 6.3 (Research Systems, Boulder, CO). Each recording to an odor stimulus corresponded to a four-dimensional array with the excitation wavelength (340 or 380 nm), two spatial dimensions ($x$, $y$ pixels of the area of interest), and the temporal dimension (100 frames). Three steps were carried out to calculate the signals (Galizia and Vetter 2005). First, to correct for bleaching at each wavelength (340 and 380 nm), a logarithmic decline curve fitted to mean fluorescence decay was subtracted from the whole image. The 5 s following odor onset were not included in the fitting process. Such a correction stabilizes the baseline of the recordings, without affecting odor-evoked responses. Second, we performed at each wavelength (340 and 380 nm) a scattered light correction, avoiding that glomeruli with strong activity cause fictive activity in neighboring silent glomeruli. We then calculated $F_n' = F_n + 3[F_n - \text{sm}(F_n)]$, where $F_n$ corresponds to frame number $n$ and $\text{sm}(F_n)$ represents $F_{n-1}$ after treatment with a boxcar filter (kernel size 50 $\mu$m, i.e., 25 pixels). Third, for each pixel, we calculated the ratio $R$ of the 340 and the 380 nm data. Then, $\Delta R_n = R_n - R_0$ was calculated, in which $R_0$ is the ratio data at frame 0 and $R_n$ is the average of a few frames before the stimulus (frames 5–13). Thus $\Delta R$ is close to 0 shortly before the odor stimulus and shows throughout a recording the relative variations of the ratio. For quantitative analysis of the data, the time course of $\Delta R$ was calculated by averaging 49 pixels ($7 \times 7$) at the center of each identified glomerulus and well within its borders. Then, amplitude of the odor-induced response of each glomerulus was calculated by subtracting the mean of 3 frames during the odor (frames 17–19) from the mean of 3 frames before the stimulus (frames 12–14). This time window corresponded

**Mapping of glomeruli**

Visualization of the glomerular AL structure after functional optical imaging was obtained according to Sandoz (2006). The brain was soon treated with a protease and 4% neutral red in distilled water. The preparation was alternately washed with Ringer (in mM: NaCl, 130; KCl, 6; CaCl$_2$, 5; MgCl$_2$, 4; sucrose, 160; glucose, 25; HEPES, 10; pH 6.7, 500 mOsmol; all chemicals were from Sigma-Aldrich, Lyon, France) and before staining, illumination was reduced and switched to red light. In this work we specifically stained according to the protocol proposed by Sachse and Galizia (2002) projection neurons conveying odor information through the lateral antennocerebral tract (LACT) from the AL to the mushroom bodies and the lateral horn using fura-2 dextran (10,000 MW, in 2% BSA; Molecular Probes). After staining, the brain was thoroughly washed with saline and the bee was left in a dark place for 3 h.

**Output (PNs)**

**Inhibition (LNs)**

**Input (ORNs)**

**Output (PNs)**

**Inhibition (LNs)**

**Input (ORNs)**

**FIG. 1.** Schematic view of the antennal lobe (AL) and odor-induced calcium activity from a population of second-order neurons (projection neurons [PNs]) in the honeybee AL. A: schematic view of the AL showing its glomerular activity and including the incoming (olfactory receptor neurons [ORNs], black) and outgoing (PNs, green) information pathways, as well as the local network of inhibitory interneurons (LNs, blue). B: the activity patterns present the amplitude of the calcium response obtained in one bee to 4 individual odorants and all the possible binary, ternary, and quaternary mixtures. The activity patterns are represented according to a common intensity scale ($F_{340}/F_{380}$ ratio, top right). In all cases (except the air control), combinations of glomeruli were activated to the individual odorants and their mixtures.
to the moment of maximal separation power between odor response and baseline (see Supplemental Fig. S6). This value was then used in all computations. For the presentation of activity maps (Fig. 1), the amplitude of odor-induced responses within each pixel was calculated as before. The resulting image was then subjected to a boxcar filter of kernel size 9 pixels and presented in a false color code of increasing amplitude from dark blue to red.

Data analysis

The goal of the present study was to compare the rules underlying olfactory-mixture representation between the input (as measured from the compound signal using bath-application of Calcium Green-2 AM; Deisig et al. 2006) and the output (measured by specific staining of the projection neurons with fura-2 dextran; this study). We first used an analysis rationale similar to that in the initial study, analyzing the relationships between mixture- and component-induced signals, both at the level of individual glomeruli and considering the whole glomerular pattern accessible to us. We also provided a range of novel analyses comparing the linearity of mixture representation between input and output and the separation power between odor representations achieved at these two levels.

NUMBER OF ACTIVE GLOMERULI. We recorded dendritic calcium signals from i-ACT PNs in AL glomeruli. Previous experiments have shown that these glomerular calcium responses correlate closely with the action potential activity of these PNs (Galizia and Kimmerle 2004). Thus calcium signals from PN dendrites are a monitor for the activity of PNs at the output side of the glomerulus (Szyszka et al. 2005). We asked how the magnitude of PN activity changes with added mixture components. We thus measured the number of glomeruli in which the recorded dendritic PN signal was above noise level (see following text), later termed “active glomeruli” for easier reference. Noise was defined as 4SD of the signal within each glomerulus before the stimulus (frames 1–14). The proportion of excitatory and inhibitory signals was determined from signals respectively above +4SD or below −4SD. Since inhibitory responses were very few, analysis concentrated on excitatory responses. The number of activated glomeruli was compared among mixture types (single odor, binary, ternary, and quaternary mixtures) or among single odors, using repeated-measure ANOVAs whenever possible (equal observation numbers).

MIXTURE INTERACTIONS. For evaluation of possible mixture interactions at the level of single glomeruli, the response classification method was the same as that used in Deisig et al. (2006). For each glomerulus the response to a mixture and the response to the strongest component (elemental model), the relative Euclidean distances between a component and the mixture in the putative olfactory space (for a calculation example, see Fig. 7A in Deisig et al. 2006). Thus for binary, ternary, and quaternary mixtures, equidistance between a mixture and its components corresponds to relative distances $D_{mix} = 50, 33.3, and 25\%$, respectively. To provide a measure of relative distance that is independent of component number ($p$), the relative distance index was calculated as $D_{mix}^r = p 	imes D_{mix}$. Thus in all cases, equidistance corresponded to a value of 1. For each mixture component, its divergence from equidistance was calculated as the absolute value of $D_{mix}^r - 1$. To evaluate how much odor representation differed from equidistance at the two processing levels, these divergence values were compared between PNs and compound signal data sets using a paired $t$-test (Fig. 4B).

To test whether the representation of a mixture follows those of the components (elemental model), the relative Euclidean distances between mixture and each of its components were compared with the relative “weight” of the components in the mixture $W_{f,mix}$. Such weight is given by the magnitude of the response (number of activated glomeruli) to the component when presented alone, relative to the sum of response magnitudes of all components of the mixture. Thus the relative “weight” ($W$) of a component in a mixture was

$$W_{f,mix} = \frac{N_i}{\sum_{k=1}^{p} N_i}$$

where $i$ indicates a component, $mix$ is the mixture, $p$ is the number of components in the mixture, and $N_i$ is the number of active glomeruli when presenting odor $k$ alone (for a calculation example, see Fig. 7B in Deisig et al. 2006). To provide a relative weight that is independent of the number of components, we calculated the relative weight index as $W_{i,mix} = p \times W_{i,mix}$. Thus for all mixtures, components having an equal weight would yield $W_{i,mix} = 1$. We then determined the

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1 The online version of this article contains supplemental data.
relationship between $D'_{\text{mix}}$ and $W'_{\text{mix}}$, which gives an overall evaluation of the quality of the model, independently of the number of mixture components. All linear correlations were tested for significance by calculating Pearson’s $r$ and using a t-test (Zar 1999; Fig. 4C). To compare the efficacy of the correlation obtained for AL input (compound signal) and output (PNs), we compared correlation coefficients using a homogeneity t-test.

**SEPARABILITY OF ODOR STIMULI.** Because PN and compound signals had different amplitudes, the Euclidean distances between the two data sets could not be directly compared. We thus used two different methods to compare separability at AL input and output levels. First, we calculated Euclidean distances on normalized data sets—i.e., by normalizing responses in all glomeruli to all odors according to the highest glomerular response observed in this data set (Fig. 5). Second, we used Pearson’s correlation coefficient $r$ as a measure of similarity between odor representations (Supplemental Fig. S5), which is independent of response amplitude (Peel et al. 2006; Sachse et al. 1999, 2002). Both methods gave the same results, thus underlining the robustness of our data.

**RESULTS**

**Calcium signals from projection neurons**

Using the calcium sensitive dye fura-2 dextran, we performed recordings from a population of uniglomerular projection neurons belonging to the lateral-antennal glomerular tract (l-ACT), which conveys olfactory information from the AL to higher-order centers such as the mushroom bodies and the lateral horn. Bees were stimulated with four single odors and all their binary, ternary, and quaternary mixtures. All stimuli induced activity in a combination of glomeruli, whereas air stimulation had no effect (Fig. 1B, example for one bee). Signals were mostly excitatory (98.8%, phasic or phasic- tonic; e.g., Fig. 2A), showing an increase in calcium concentration on stimulation, which is in accordance with previous PN recordings in the honeybee (Peel et al. 2006; Szyszka et al. 2005).

**Number of activated glomeruli**

When considering the intensity of glomerular activation on single odorant and mixture stimulation, we found output response properties similar to those previously recorded at AL input (compound signal; Deisig et al. 2006). First, the four primary odorants yielded a significant heterogeneity in glomerular activation when PN responses were quantified [repeated-measures ANOVA, $F_{(3,24)} = 5.88$, $P < 0.004$; Supplemental Fig. S1A], as at the input (Deisig et al. 2006). In both data sets, the primary odors 1-hexanol and 2-octanol induced stronger activation ($8.3 \pm 1.7$ and $7.9 \pm 1.2$ activated glomeruli, respectively: means $\pm$ SE), whereas linalool and limonene induced weaker activation ($5.4 \pm 1.2$ and $3.8 \pm 0.7$ activated glomeruli, respectively). Second, in both data sets, the number of activated glomeruli strongly saturated with an increasing number of components in the mixture. In the case of PNs, no significant difference was found between stimuli with an in-
creasing number of components [Supplemental Fig. S1B: ANOVA, \( F_{(3,131)} = 0.19, \) NS]. Third, response magnitudes to the 15 odor stimuli were significantly correlated between AL input and output (Supplemental Fig. S2C, \( r = 0.70, P < 0.01, \) degrees of freedom [df] = 13). Fourth, for all odors and all glomeruli, the amplitudes of calcium responses were highly correlated between the two data sets (data not shown; \( r = 0.59, P < 0.001, \) df = 312). A significant correlation was found between PN and compound signal activity patterns in 12 of 15 cases \( (r > 0.43, P < 0.05, \) df = 19). This correlation was significantly higher for primary and binary olfactory stimulations \( (r = 0.60 \pm 0.03) \) than that for ternary and quaternary ones \( (r = 0.47 \pm 0.04; t = 2.67, P = 0.02, \) df = 13). These data show that AL local processing maintains similar quantitative relationships between the four single odorants or between mixtures with increasing numbers of components.

**Mixture interactions at the glomerular level**

Average PN activity maps were constructed for the nine bees tested (Supplemental Fig. S2). Although AL output activity patterns were qualitatively similar to AL input patterns, some glomeruli were exclusively activated in the compound signal data and others rather exclusively in the PN data. As at the input, a glomerulus exhibited PN activity on mixture stimulation when it was activated by at least one of its components. However, directly comparing the amplitudes of mixture and component responses revealed mixture responses that could not be easily predicted from the response to the components. PN responses were thus classified according to categories defined by Duchamp-Viret et al. (2003) (for details see Deisig et al. 2006). Most glomeruli showed suppression (lower response to the mixture than that to the most effective component; Fig. 2A, top) and hypoadditivity (equal responses to the mixture and to the most effective component; Fig. 2A, bottom), whereas cases of synergy (higher response to the mixture than that to the most effective component; not shown) were rare. We found that increasing the number of mixture components significantly increased suppression [Supplemental Fig. S3, ANOVA, \( F_{(2,96)} = 5.77, P < 0.005, \) whereas the number of hypoadditivity and of synergy cases did not change [respectively, \( F_{(2,96)} = 0.40, \) NS and \( F_{(2,96)} = 0.21, \) NS]. A substantial difference appeared between AL input and output at this level (Fig. 2B): PN signals showed significantly more suppression than that shown by compound signals [population × component number ANOVA, population effect: \( F_{(1,203)} = 6.11, P = 0.014; \) component number effect: \( F_{(2,203)} = 16.9, P < 0.001, \) and logically less hypoadditivity [population effect: \( F_{(1,203)} = 13.6, P < 0.001; \) component number effect: \( F_{(2,203)} = 0.19, \) NS]. This indicates that local networks in the AL produce increased suppression of mixture activity at AL output relative to its input.

**Similarity between mixture and single odor representations**

We evaluated similarity relationships between odor representations on the basis of Euclidean distances within a putative olfactory space, in which each dimension is represented by activity within one of the recorded glomeruli. Similarity between mixture and component representations was globally conserved from AL input to output (Fig. 3A; correlation of distances between all possible odor pairs from both data sets, \( r = 0.66, P < 0.001, \) df = 103). A PCA revealed three very conspicuous groups of odors, based on their similarity relationships (Fig. 3B, the first two factors represented 66.2% of overall variance). This picture was confirmed by a cluster analysis (Fig. 3C; see PN column). Cluster 1 grouped the two weaker odors (in terms of the number of active glomeruli; see Supplemental Fig. S1A) limonene, linalool, and their binary mixture. Cluster 2 included the strong odor 1-hexanol and all its mixtures with linalool and/or limonene. Cluster 3 grouped the strong odor 2-octanol and all its mixtures and showed a subdivision. One subcluster included all mixtures containing 2-octanol and limonene; the other subcluster contained 2-octanol and all its mixtures with 1-hexanol and linalool. This repartition strongly correlated with that found for the input (Deisig et al. 2006 and Fig. 3C; see compound signal column). Thus although more suppression is evident at the output of the AL, local processing does not modify mixture representation in a drastic way.

**Linearity of mixture representations**

Similarity relationships among odors found within both the input and the output neural spaces were correlated, but whereas compound signal-based similarity relationships between a mixture and its components could be predicted by an elemental model based on the physiological salience of components (defined as the number of glomeruli activated by each component), a strong and significant departure from the elemental model was observed at the PN level. This is illustrated by the relative Euclidean distances between the representations of each mixture and its components, when calculated in both the glomerular multidimensional input (compound signal, in black) and the output (PN, in white) spaces (Fig. 4A, representative example of the quaternary mixture; Supplemental Fig. S4 shows the same comparison for all mixtures). Bars of equal length correspond to a perfect balance (equidistance) between the components of a mixture (50% for binary, 33% for ternary, and 25% for the quaternary mixture; dotted lines in Fig. 4A and Supplemental Fig. S4). Although compound signal data revealed strongly asymmetrical representations of components in the mixtures (which were explained by different physiological saliences of the components; see following text), component representations in the PN data were more equidistant to their mixture. Components were thus more balanced within mixture representation at the PN level in four of the six binary mixtures, in all ternary and in the quaternary mixture. When analyzing the mean divergence of each component to equidistance in the two data sets (Fig. 4B, 0.29 ± 0.03 for compound signal and 0.17 ± 0.02 for PN), components were significantly closer to equidistance at the output level than at the input level (paired \( t \)-test, df = 26, \( P = 0.0012 \)). As a consequence (Fig. 4C), the elemental principle holding for the input is the stronger a component is (i.e., the more glomeruli it activates), the more similar the mixture is to this component (i.e., the smaller the distance between mixture and component). Indeed, we found a highly linear negative correlation coefficient between these two variables \( (r = \) -0.94). By contrast, output representation showed a much more scattered trend, with a still negative but lower correlation coefficient \( (r = \) -0.60). Although both correlations were significant \( (P < 0.001, 26 \) df),
the coefficient for PNs was significantly lower than that obtained for compound signals (homogeneity \( t \)-test, \( P < 0.001 \)). Thus when compared with the AL input, output mixture representations display a significant departure from the elemental model, which may be attributed to local processing and increased suppression as found before.

Separability between odor representations

What could be the benefit of such fine modifications of mixture representation in the AL? We hypothesized that such reformatting may improve odor separability. To test this idea, we compared Euclidean distances between glomerular activation maps based on normalized input and output data sets. In line with our hypothesis, odor response patterns were significantly more dissimilar, i.e., distances were higher for PNs than those for the compound signal over all possible odor pairs (\( n = 105 \), Fig. 5A, Wilcoxon matched-pair test, \( z = 3.06, P = 0.002, df = 104 \)). We also found that distances were significantly higher at the PN level for comparisons between any two of the mixtures (Fig. 5B, \( n = 55, z = 2.49, P = 0.023 \)), but not when comparing mixtures to single odors (“elements,” Fig. 5B, \( n = 44, z = 1.30, NS \)). Comparisons between single odors were also not significant (data not shown, \( n = 6, z = 1.6, NS \)). Thus AL processing increased Euclidean distances between mixture representations. This effect was not due to data normalization because distances between odors and the air control were identical for the compound signal and PN data sets, after normalization (1.78 ± 0.10 and 1.79 ± 0.09, respectively, \( z = 0.17, NS \)). Moreover, this effect was confirmed by using correlation coefficients between response patterns, a measure of pattern similarity that is independent of differences in the two datasets. This analysis showed again that output activity patterns were less correlated than input patterns (\( n = 105 \), Supplemental Fig. S5, \( z = 2.37, P = 0.017 \)). This effect appeared for comparisons among mixtures (\( z = 5.32, P < 0.001 \)), but was not significant for comparisons between mixtures and single odors (\( z = 1.61, NS \)). Thus the main effect of AL processing was to decorrelate mixture activity patterns.

Time course of component–mixture relationship

As shown earlier, projection neurons can show polymorphic responses, including phasic, phasic-tonic, and sometimes more complex time courses after stimulus presentation (Sachse and Galizia 2002). We thus asked whether the relationship between mixture and component representations changes with time and thus whether the rules we describe apply throughout the epoch of projection neuron activity. To do this, we computed Euclidean distances between mixture and component representations at each time point, after setting all data to 0 at the first frame (to avoid calculation artifacts before the stimulus). We represented the evolution of the average distance between each mixture type and its components throughout an entire recording (Supplemental Fig. S6). At odor onset, dis-
tances between stimuli increased dramatically on stimulus delivery and reached a maximum about 600 ms after stimulus onset, decreasing to near baseline after stimulus offset. The evolution of the distance was the same for all mixture types (binary, ternary, quaternary) and closely followed the evolution of the distance between single odors. When evaluating the correlation between the different curves at signal maximum (2 s during which the distance is at its highest, i.e., 200 to 2,200 ms after stimulus onset; see dotted line in Supplemental Fig. S6), all coefficients were >0.991. This indicates that the relationships between mixture and component representations follow the same time course throughout a recording.

**DISCUSSION**

Our study shows that AL processing introduces fine rearrangements in mixture neural representations, so that 1) components are more homogeneously represented within a mixture representation at the PN level than at the input level, 2) mixture representation is less predictable by an elemental model, and 3) odor mixture representations become more separable after AL processing.

Recent works investigated reformatting of odor representation within the AL and compared input and output representations of a range of monomolecular odorants. These studies, performed with different techniques, revealed contrasting processing principles, but led to a similar conclusion. Optical imaging in honeybees illustrated that AL processing sharpens odor response profiles of PNs, leading to better differentiation between single odorants (Sachse et al. 2002, 2003). By contrast, electrophysiology in fruit flies revealed a broadening of odor response profiles of PNs but, as in bees, this modification led to an improved discrimination between odorants (Bhandawat et al. 2007). Because olfactory mixtures constitute more natural stimuli than monomolecular odors we studied AL-induced modifications in mixture representation. We show that a rather fine rearrangement occurs from AL input to output and that such a rearrangement results in a higher separation power between complex odors. Several studies suggested that mixture representation is rather elemental at the input level (zebrafish: Tabor et al. 2004; bees: Deisig et al. 2006; moths: Carlsson et al. 2007; fruit flies: Silbering et al. 2007), even though within individual ORNs nonlinear responses to mixtures can be observed (Akers and Getz 1993; Cromarty and Derby 1998; Duchamp-Viret et al. 2003). At the output level, studies in the zebrafish (Tabor et al. 2004) and in *Drosophila* (Silbering et al. 2007) showed an increased proportion of cases of nonlinear summation between component signals within individual neurons or glomeruli. Increased suppression phenomena at the PN level in our study fully confirm these accounts (Fig. 2B). However, there is an important choice to make when comparing two processing levels, that is 1) focusing on particular units (as in most studies) or 2) describing odor representation in its multidimensional (global) quality (Bhandawat et al. 2007; Broome et al. 2006; this study). Both approaches are certainly helpful; however, the whole odor response pattern seems to...
have more consequence on the animal's behavior than individual units, in that the latter allows predicting olfactory behavior (Guerrieri et al. 2005). Previous studies neither analyzed the multidimensional quality of mixture representation nor did they evaluate the separability of mixture representations at both processing levels. Our study is the first to evaluate these points.

For comparing mixture representation at two processing levels, we used in vivo calcium imaging based on two different staining techniques. As with any experimental approach, some cautionary arguments should be made, which relate to 1) the cell populations participating in the signals and 2) the link between calcium signal and neural activity.

CELL TYPES. For the input signal, we bath-applied a permeable calcium sensitive dye, Calcium Green-2 AM, on the brain. With this protocol, all AL cells could be potentially stained (ORNs, PNs, local inhibitory interneurons, glia), but ORNs are by far the most numerous. In addition, the recorded signals are highly stereotyped and never show any spontaneous activity or any inhibitory responses, which are typical for local neurons and PNs (Sachse and Galizia 2002). Therefore the participation of LN or PN s in the compound signal is thought to be negligible (Galizia and Vetter 2005). A significant part of the signal may come from glial cells surrounding each glomerulus because in the hawkmoth Manduca sexta, bath application of Calcium Green was shown to mainly stain periglomerular glial cells in adults and odors trigger activity in these cells (Heil et al. 2007). Even so, it is clearly established that release of acetylcholine (ACh) from ORN axons leads to activation of nicotinic receptors (nAChRs) in these glial cells, which depolarizes the cell membrane and thereby opens voltage-gated calcium channels (VGCCs). In other words, even if glial cells participate in the calcium signal, they do so in a manner directly correlated to ORN input. We thus interpret the compound signal as mainly representing sensory input. For recording the output representation (present recordings), we used retrograde staining of PNs with fura-2 dextran and the signal was recorded from the glomeruli (i.e., the dendrites of these neurons in the AL). In this case, there is no doubt that only PNs participated in the signal because the dye was injected rather far away from the AL and confocal scans confirmed the specificity of the staining (Sachse et al. 2002; our observation).

CALCULUM SIGNAL AND NEURAL ACTIVITY. The second question relates to the quantitative relationship between the recorded calcium signals and spiking activity of olfactory neurons. Calcium imaging is used as a standard technique for recording neural activity at the network level in many systems (Kwan 2008), since it has been consistently shown that the recorded fluorescence changes are a direct result of calcium influx from action potentials (Kerr et al. 2005) and can be used to reconstruct firing rates efficiently (Yaksi et al. 2006). In our system several arguments indicate a tight link between firing rate and calcium signal. First, recordings of both electrophysiological activity and calcium imaging showed that excitatory (respectively, inhibitory) electrical activity is accompanied by positive (respectively, negative) calcium signals in AL neurons (Galizia and Kimmerle 2004). In addition, calcium imaging of honeybee PNs in primary cell cultures and pharmacological applications performed in our laboratory showed that ACh, which depolarizes the membrane and triggers spiking activity, induces dose-dependent calcium signals predominantly (>80%) through VGCC (V. Raymond-Delpech, M. Augier, T. Jaillard, J. C. Sandoz, unpublished data). Therefore we believe that the calcium signals recorded in our system are closely related to the electrical activity of the neurons.

FIG. 5. Antennal lobe processing increases separability between odor mixtures. A: Euclidean distances (means ± SE) were calculated on normalized input (compound signal, black) and output (PN, white) data sets. At the PN level, odors were more separable than at the input level because the mean Euclidean distance between any two odors was significantly higher (Wilcoxon matched-pair test, P < 0.01; 104 df). B: mean Euclidean distances are shown for comparisons involving (left) any mixture and a single odor ("element," n = 44 possibilities) and (right) any 2 mixtures (n = 55 possibilities). Distances were significantly higher at the PN level than at the input level for comparisons involving 2 mixtures (Wilcoxon test, P < 0.01). A similar analysis based on correlation coefficients is given in Supplemental Fig. S5. Overall, odor mixture representations are more readily separable at the AL output than at the input.
With these comments in mind, our study shows that AL processing globally conserves the overall structure of the odor space (Fig. 3B) and keeps a similar coarse classification of odor representations (Supplemental Fig. S5). However, AL processing improves fine odor-similarity relationships. A careful evaluation of mixture–component relationships shows that components are more homogeneously represented at the PN level than at the input level (Fig. 4B; see following text). This work also shows for the first time that such interactions bestow improved separability of mixture representations. Both observations suggest that, as in fruit flies (Bhandawat et al. 2007), AL processing produces more uniform distances between odor representations, thereby increasing the global efficiency of the system for odor discrimination (Fig. 5A). In other words, PNs appear to use their multidimensional coding space better than ORNs, both for single odors (Bhandawat et al. 2007) and for odor mixtures (this study). Components were more homogeneously represented within mixture patterns at the PN level (Fig. 4B). This may be a result of the decorrelation of odor representations through AL processing. If AL processing provokes more homogeneously distributed representations in the PN space, it could have the collateral effect of decreasing asymmetries in component–mixture relationships. A direct consequence would be that mixture representation would no longer contain precise information about the relative quantity of its components. If so, PN mixture representation should be more concentration invariant with regard to differences in components’ quantity than at the ORN level. Such a property could be important for honeybees, which are “flower constant,” thus limiting their visits to a single plant species as long as it is profitable (Grant 1950). Floral aromas are multicomponent blends that fluctuate in composition according to genotype and flowering stage (Pham-Delégue et al. 1989). Our results indicate that the olfactory system may provide such a component-concentration invariant representation.

Our results fully support a two-tiered model of lateral inhibition in the AL (Sachse and Galizia 2002; Silbering and Galizia 2007), with one tier representing homogeneous inhibition, playing the role of a gain control and a second tier representing heterogeneous, asymmetrical inhibition between glomeruli. We observed the first tier at the input level because inhibition reduced overall signal strength, thereby avoiding saturating the system, but kept elemental information within mixture representation (Deisig et al. 2006). PN results point to the second tier, with an inhibitory system provoking more interactions between components within mixture signals and modifying fine odor-similarity relationships. Such a system could explain the observed departure from elemental mixture representation. What could be the neural basis of such a model? The honeybee AL contains two main anatomical types of local inhibitory interneurons, possibly representing the two tiers: roughly 13% of the 4,000 local interneurons diffuse homogeneously in the AL (homo-LNs), whereas the rest show a high branching density in one particular glomerulus (hetero-LNs: Flanagan and Mercer 1989; Fonta et al. 1993). HomolNs would be involved in gain control (first tier) and are thought to represent a picrotoxin-sensitive network, based on GABAergic neurotransmission (Sachse and Galizia 2002). On the other hand, hetero-LNs would accomplish focused lateral inhibition between physiologically neighbor glomeruli (second tier) and would constitute a picrotoxin-insensitive network (Sachse and Galizia 2002). The neurotransmitter for this second tier is still unknown. Recently, a histaminergic inhibitory system has been proposed (Sachse et al. 2006), but it likely belongs to the first tier because histamine-imunoreactive interneurons are few (∼35 LNs; Bornhauser and Meyer 1997) and histamine application during calcium imaging already provokes a strong reduction of odor-evoked signals at the input level (Sachse et al. 2006). Other candidates could be glutamate (Barbara et al. 2005; El Hassani et al. 2008), but also γ-aminobutyric acid through possible picrotoxin-insensitive heteromeric chloride channels (Dupuis et al. 2010). In contrast to Drosophila, up to now, no excitatory interneurons have been found in the bee.

The honeybee olfactory system is mainly organized along two parallel pathways, with two main types of uniglomerular PNs taking information from different subsets of AL glomeruli (Abel et al. 2001; Kirchner et al. 2006; Müller et al. 2002). Comparing the rules described here for lateral-AL neurons with those of the second population (median-AL) could be especially promising: 1-AL neurons could provide the brain with component-composition invariant information, whereas m-AL neurons would keep track of relative component quantities. A necessary step should thus be the study of the subsequent reformatting of mixture representations in higher-order brain centers. The strong sparsening of odor representation from PNs to Kenyon cells, the intrinsic mushroom body neurons (Perez-Orive et al. 2002; Szyzka et al. 2005), and their coincidence detection properties (Fortner et al. 2007) could be the basis for mixture-specific units, as in the mouse olfactory cortex. There, about 30% of all recorded neurons were mixture specific, responding to binary mixtures, but not to either of their components (Zou and Buck 2006). Whether equivalent units exist in the mushroom bodies remains to be tested.

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DISCLOSURES

No conflicts of interest are declared by the authors.

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